

## DITERPENOID CONSTITUENTS FROM *RABDOSIA WEISIENSIS*

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**Key Word Index**—*Rabdosia weisiensis*; Labiatae; trichorabdal A; *ent*-1 $\beta$ ,7 $\alpha$ ,11 $\alpha$ -triacetox-3 $\alpha$ ,6 $\beta$ -dihydroxy-kaur-16-en-15-one.

**Abstract**—A new diterpenoid was isolated from the ethereal extract of the dried leaves of *Rabdosia weisiensis* and its chemical structure shown to be *ent*-1 $\beta$ ,7 $\alpha$ ,11 $\alpha$ -triacetox-3 $\alpha$ ,6 $\beta$ -dihydroxykaur-16-en-15-one on the basis of detailed spectroscopic analysis and comparison with related compounds. Trichorabdal A, a known component, was also isolated from this plant and its  $^{13}\text{C}$  NMR data assigned.

*Rabdosia weisiensis* C. Y. Wu is distributed over the northwestern district of Yunnan, China [1]. An ethereal extract of its dried leaves was fractionated by column chromatography (CC) on silica gel. Components were further purified by numerous silica gel CC and/or silica gel preparative TLC and crystallization.

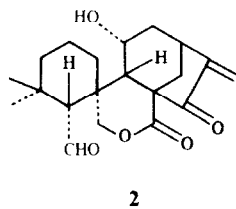
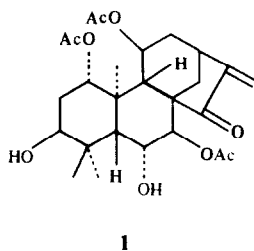
Weisiensin A (**1**),  $\text{C}_{26}\text{H}_{36}\text{O}_9$ , mp 298–300°, showed the presence of three methyl groups, three methylene groups, eight methine groups, three quaternary carbons, two olefinic carbons, one ketonic carbon and three acetoxy groups in the  $^{13}\text{C}$  NMR (DEPT) spectrum (Table 1). It had a five-membered ketone conjugated with an *exo*-methylene group, judging from the following spectral data:  $\lambda_{\text{max}}^{\text{EtOH}}$  236.5 nm (log  $\epsilon$  3.86);  $\nu_{\text{max}}^{\text{KBr}}$  1740–1708, 1645  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR:  $\delta$  5.91 and 5.13 (each 1H, *br s*),  $^{13}\text{C}$  NMR:  $\delta$  111.72 (*t*), 151.21 (*s*) (*exo*-methylene) and 205.51 (ketone) [2]. The above-mentioned data together with three tertiary methyl signals at  $\delta$  1.74, 1.57 and 1.30 in the  $^1\text{H}$  NMR spectrum suggested that this compound had the typical *ent*-kaur-16-en-15-one nucleus as a basic skeleton [3]. Weisiensin A was found to have two hydroxy groups and three acetoxy groups:  $\nu_{\text{max}}^{\text{KBr}}$  3570, 3520, 1740–1708, 1252  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR:  $\delta$  6.08 (1H, *d*,  $J$  = 3.96 Hz), 5.83 (1H, *dd*,  $J$  = 10.08, 5.76 Hz), 5.58 (1H, *d*,  $J$  = 3.60 Hz), 4.47 (1H, *br s*,  $W_{1/2}$  = 5.0 Hz), 3.71 (1H, *t*,  $J$  = 3.0 Hz), 2.21, 2.14 and 1.967 (each 3H, *s*).  $^{13}\text{C}$  NMR:  $\delta$  81.01, 76.26, 75.81, 70.40, and 69.38 (each *d*).

The locations of five oxygen functional groups were deduced as follows. The chemical shift values of C-10 and C-20 were  $\delta$  43.62 and 16.06, respectively, which suggested that there was an oxygen substituent on the 1 $\alpha$ -position [4]. In the  $^{13}\text{C}$  NMR spectrum C-4 was at  $\delta$  38.84, C-18 at 29.53 and C-5 at 41.66 which suggested that there were

oxygen substituents on the 3 $\beta$ - and 7 $\beta$ -positions [5, 6]. The chemical shift value of C-5 was  $\delta$  41.66 and C-19 was 24.68 which indicated that there was an oxygen substitute on the 6 $\alpha$ -position [5, 6]. The C-13 signal was at  $\delta$  37.10 which indicated there were no oxygen substituents on C-12 or C-14 [7]. The C-9 signal was at  $\delta$  55.62 and C-8 was at 49.14 which suggested that there was an oxygen substitute on the 11 $\beta$ -position [6]. The functional group at C-3 $\beta$  was a hydroxyl judged by  $^1\text{H}$  NMR data ( $\delta$  3.71, *t*,

Table 1.  $^{13}\text{C}$  NMR chemical shifts of weisiensin A (**1**) and trichorabdal A (**2**)

C	1	2
1	81.01 <i>d</i>	31.60 <i>t</i>
2	33.37 <i>t</i>	18.64 <i>t</i>
3	75.81 <i>d</i>	40.16 <i>t</i>
4	38.84 <i>s</i>	34.31 <i>s</i>
5	41.66 <i>d</i>	56.91 <i>d</i>
6	69.38 <i>d</i>	205.41 <i>d</i>
7	76.26 <i>d</i>	171.26 <i>s</i>
8	49.14 <i>s</i>	60.70 <i>s</i>
9	55.62 <i>d</i>	47.70 <i>d</i>
10	43.62 <i>s</i>	42.06 <i>s</i>
11	70.40 <i>d</i>	64.98 <i>d</i>
12	38.64 <i>t</i>	42.65 <i>t</i>
13	37.10 <i>d</i>	35.23 <i>d</i>
14	36.84 <i>t</i>	28.51 <i>t</i>
15	205.51 <i>s</i>	201.62 <i>s</i>
16	151.21 <i>s</i>	150.24 <i>s</i>
17	111.72 <i>t</i>	117.66 <i>t</i>
18	29.53 <i>q</i>	32.30 <i>q</i>
19	24.68 <i>q</i>	26.12 <i>q</i>
20	16.06 <i>q</i>	70.94 <i>t</i>
OAc	170.29 <i>s</i>	
	170.22 <i>s</i>	
	169.26 <i>s</i>	
	21.89 <i>q</i>	
	21.37 <i>q</i>	
	21.10 <i>q</i>	



$J = 3.0$  Hz) [8]. The location of the other hydroxyl group at C-6 $\alpha$  was supported by the following decoupling experiments. On irradiation at  $\delta 4.47$  (*br s*, 6 $\beta$ -H), the doublet at 5.58 (7 $\alpha$ -H) changed into a singlet and the broad singlet at 2.66 (5 $\beta$ -H) changed into a sharp singlet. On the other hand, on irradiation at  $\delta 5.58$ , the broad singlet at 4.47 sharpened. Further, on irradiation at 6.08 (11 $\alpha$ -H), the broad singlet at 2.76 (9 $\beta$ -H) collapsed to a sharp singlet and the sextet at 2.36 (12 $\beta$ -H,  $J = 2.0$ , 2.0, 14.0 Hz) changed into a double doublet ( $J = 2$ , 0, 14.0 Hz). Therefore, the chemical structure of weisiensin A (1) could be represented as *ent*-1 $\beta$ ,7 $\alpha$ ,11 $\alpha$ -triacetoxy-3 $\alpha$ ,6 $\beta$ -dihydroxykaur-16-en-15-one (1). Trichorabdal A (2), a known component [9], was also isolated from this plant and its  $^{13}\text{C}$  NMR data are assigned in Table 1.

#### EXPERIMENTAL

The experimental conditions were the same as those described in the previous paper [10].

**Plant material.** *Rabdosia weisiensis* C. Y. Wu leaves were collected by Prof. Liu Lunhui in Weixi, Yunnan, China in Sept. 1983 and the voucher specimen been deposited in our Institute.

**Extraction and isolation of constituents.** Dried and powdered leaves (0.75 kg) were worked-up in the same manner as the previous paper [10] yielding 1 (350 mg) and 2 (105 mg).

**Weisiensin A (1).**  $\text{C}_{26}\text{H}_{36}\text{O}_9$  ( $\text{M}^+$  at  $m/z$  492), colourless needles, mp 298–300°; UV  $\lambda_{\text{max}}^{\text{EtOH}}$  nm: 236.5 (log  $\epsilon$  3.86); IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3570, 3520, 1740–1708, 1645, 1252, 1025, 935; MS  $m/z$ : 492 [ $\text{M}$ ] $^+$ , 432, 390, 372, 330, 312, 294, 279, 43 (base peak).  $^1\text{H}$  NMR:  $\delta$  6.08 (*d*,  $J = 3.96$  Hz, 11 $\alpha$ -H), 5.91 and 5.13 (each 1H, *br s*, 17-H $_2$ ), 5.83 (*dd*,  $J = 10.08$ , 5.76 Hz, 1 $\beta$ -H), 5.58 (*d*,  $J = 3.60$  Hz, 7 $\alpha$ -H), 4.47 (*br s*,  $W_{1/2} = 5.0$  Hz, 6 $\beta$ -H), 3.71 (*t*,  $J = 3.0$  Hz, 3 $\alpha$ -H), 3.20 (*d*,  $J = 12.80$  Hz, 14 $\alpha$ -H), 2.94 (*m*,  $W_{1/2}$

$= 7.0$  Hz, 13 $\alpha$ -H), 2.76 (*br s*, 9 $\beta$ -H), 2.66 (*br s*, 5 $\beta$ -H), 2.36 (*sextet*,  $J = 14.0$ , 2.0, 2.0 Hz, 12 $\beta$ -H), 2.21, 2.14 and 1.97 (each 3H, *s*, 3  $\times$  OAc), 1.74 (3H, *s*, 20-Me), 1.57 (3H, *s*, 19-Me), 1.30 (3H, *s*, 18-Me).

**Trichorabdal A (2).**  $\text{C}_{20}\text{H}_{26}\text{O}_5$ ; colourless needles, mp 202–204°; UV  $\lambda_{\text{max}}^{\text{EtOH}}$  nm: 231.5 (log  $\epsilon$  3.93); IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3435, 2800, 2720, 1745, 1722, 1688, 1643, 1275, 1178, 1020, 955, 926;  $^1\text{H}$  NMR:  $\delta$  10.07 (*d*,  $J = 4.70$  Hz, CHO), 6.02 and 5.38 (each 1H, *br s*, 17-H $_2$ ), 5.29 and 5.06 (each 1H, *d*,  $J = 12.0$  Hz, 20-H $_2$ ), 4.63 (*m*,  $W_{1/2} = 10.0$  Hz, 11 $\beta$ -H), 3.50 (*d*,  $J = 11.74$  Hz, 14 $\alpha$ -H), 2.91 (*d*,  $J = 4.70$  Hz, 5 $\beta$ -H), 1.00 and 0.97 (each 3H, *s*, 2  $\times$  Me).

#### REFERENCES

1. Wu Cheng-Yih and Li Hsi-Wen, (1977) *Flora Reipublicae Popularis Sinicae* Vol. 66, p. 526. Beijing Academia Press, Beijing.
2. Xu Yunlong, Sun Xichang, Sun Handong, Lin Zhongwen and Wang Dezu. (1981) *Acta Bot. Yunn.* 3, 283.
3. Fujita, E., Nagao, Y. and Node, M. (1976) *Heterocycles*, 793.
4. Kubo, I., Miura, I., Kamikawa, T., Isobe, T. and Kubota, T. (1977) *Chem. Letters*. 1289.
5. Qin Chongqiu, Liu Chenjiang, Li Jicheng, An Xinzong, Sun Handong, Lin Zhongwen (1984) *Acta Bot. Yunn.* 6, 333.
6. Li Jicheng, Liu Chenjiang, Sun Handong, Lin Zhongwen. (1986) *Acta Bot. Yunn.* 8, 93.
7. Kubo, I., Nakanishi, K., Kamikawa, T., Isobe, T. and Kubota, T. (1977) *Chem. Letters*. 99.
8. Sun Handong, Lin Zhongwen, Fu Jian, Zheng Xinrong, Gao Zengyi. (1985) *Acta Chim. Sin.*, 43, 353.
9. Node, M., Sai, M., Fuji, K. and Fujita, E. (1982) *Chem. Letters*. 2023.
10. Xu Yunlong and Ma Yunbao (1989) *Phytochemistry* (in press).